The Incubation Handbook

Brinsea Incubation Specialists

CONTENTS

1.

| Introduction | | Page |
|--------------|--|------|
| Describ | bes the characteristics of the two types of hatched chick and the conditions | |
| require | d by eggs for successful incubation. | |
| 1.1. | Preface | 2 |
| 1.2. | Incubation Basics | 2 |
| | | |

2. The Egg

Details the stages of development of the embryo, the dietary requirements of the parent birds and correct conditions for storing eggs prior to incubation.

| 2.1. | Embryo Development 3 |
|------|--------------------------------|
| 2.2. | Nutrition of the Egg 5 |
| 2.3. | Storage and Handling of Eggs 6 |

3. Incubator Construction and Function

Describes the general form and construction of commercially available incubators and how they work. Temperature, humidity and egg turning requirements are discussed in detail

| 3.1. | Introduction | 7 |
|------|---|----|
| 3.2. | Still Air Incubators | 7 |
| 3.3. | Forced Draft Incubators | 8 |
| 3.4. | Contact Incubators | 9 |
| 3.5. | Design Requirements for Successful Hatching | 11 |

4. Monitoring of Eggs and the Incubator

Describes the techniques and calculations used to measure the development of the egg and the incubator conditions.

| 4.1. | Candling | 15 |
|------|------------------------------------|----|
| 4.2. | Measuring Humidity and Weight Loss | 16 |

5. Troubleshooting

Table of problems likely to be encountered and the possible causes of these problems. 18

6. Species Reference Table

Incubation data for commonly bred species.

21

1. Introduction

1.1. Preface

This handbook has been written to provide the reader with a knowledge of incubation principles and a practical guide to incubation techniques. It describes the process of embryo development and how incubators are designed to provide the ideal conditions for this process. The handbook deals with problems likely to be encountered and gives recommendations for overcoming them as well as useful calculation tools for monitoring the development of the egg. Although much of the scientific data is based on poultry eggs it is generally applicable to all species. Where differences exist these are explained.

1.2. Incubation basics

All bird eggs require five environmental conditions to be controlled to enable the correct development of the embryo:

- The egg must be maintained at the right temperature to enable the metabolic processes within the developing embryo to occur at the correct rate.
- The egg must be frequently turned and carefully positioned so that the embryo passes through fresh nutrients in the white of the egg, while forming in the correct position for hatching.
- The egg loses water through pores in the shell. The humidity of the air around it must be controlled to ensure the right amount of water is lost over the incubation period.
- The egg "breathes" so there must be a supply of fresh air to provide oxygen and to remove waste carbon dioxide. Eggs are susceptible to infection so the incubator must provide a clean, disinfected environment.

1.3. Precocial and Altricial Species

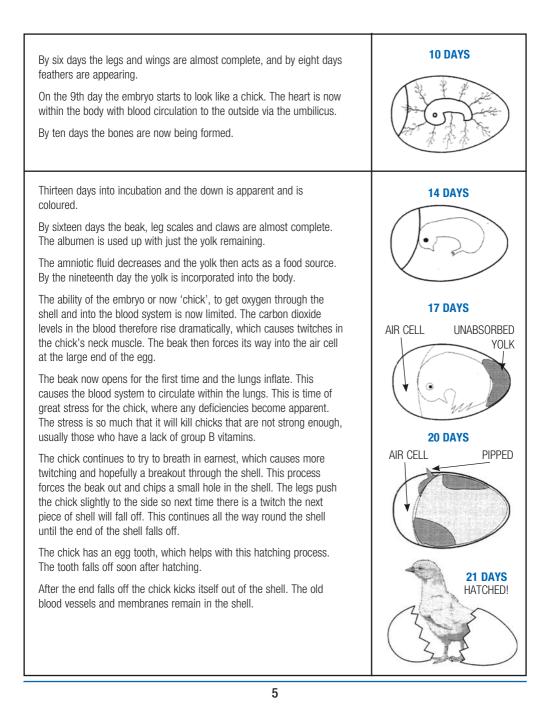
The words Precocial and Altricial describe the degree of development in young birds at hatching, precocial meaning a bird that is 'capable of moving around after hatching' and altricial meaning a bird is 'incapable of moving around after hatching'. Although the basic principles of incubation apply equally to both groups there are differences and it must be noted that specific incubation conditions should be created for the individual species concerned.

| Precocial Characteristics | Altricial Characteristics |
|---|---|
| Hatched with eyes open Covered with down Leaves nest within two days Will forage for food within hours Eggs commonly turned less frequently | Hatched with eyes closed Little or no down Incapable of departing from nest Fed by parents Eggs commonly turned more frequently |
| Example Species:- Poultry, Waterfowl | Example Species:- Birds of Prey, Parrots |

2. The Egg

2.1. Embryo Development (based on hen egg)

| The yolk is dropped from the ovary into the unfundibilum where it is fertilised by the male sperm if present. | FERTILISATION And First 24 Hours |
|--|---|
| The embryo commences development while the yolk travels down the egg canal. Here it receives coatings of white. The egg progresses to the shell gland where it receives membranes | |
| and forms the shell itself. This process takes approximately 20 hours, in which time the embryo grows to about 4mm in diameter. The egg is laid. | FERTILE EMBRYO 4mm DIA ON YOLK |
| After the egg is laid it cools and growth slows or stops. Many birds (but not all) accumulate a 'clutch' of eggs before commencing incubation. During this 'storage' time temperature is preferably between 0 and 20°C but some species tolerate more extreme temperatures. However, embryonic death is a risk if extremes are maintained. | 2 DAYS |
| In the first day of incubation a line called the primitive streak appears. This allows the formation of a third layer of cells. From these new cells the organs of the body will form. The cells are made up in three layers called ectoderm, mesoderm and endoderm. | |
| Ectoderm forms the skin, feathers, beak, nervous system, claws eyes and mouth. Mesoderm forms the skeleton, muscle, blood and reproductive organs. Endoderm forms the respiratory organs, secretary system and digestive system. | YOLK |
| By the end of the first day's incubation, the head, eyes, nervous system and blood island have started to form. The heart is formed on the second day and is functioning by 44 hours. | TULK |
| On the fourth day the heart changes from its simple form and becomes a fully formed beating heart. | 4 DAYS |
| During this time extra membranes are formed to produce the amniotic sack. The embryo will now float in the amniotic fluid for the rest of incubation. | BLOOD VEINS |
| The amniotic fluid and turning of the egg ensure the embryo orients itself correctly for hatching. | |
| By the fourth day, legs and wing buds begin to form and the heart is still positioned outside the body. | Az ip |



2.2. Nutrition of the Egg

The egg is composed of many nutrients that come from the parent so, to ensure a healthy hatched chick, the parent needs plenty of nutrients at least two weeks prior to laying. The most important vitamins and minerals are as follows:-

- Vitamin A Sources of this vitamin are in green feeds, yellow vegetables, maize and synthetic compounds. The
 vitamin can be easily destroyed by light and heat. A lack of this vitamin is a common cause of poor hatchability,
 weak chicks and poor disease resistance.
- Vitamin D The raw source of this vitamin is in sunlight. Chicks kept indoors will need supplements. A lack of it causes week bones that will bend and deform. Also eggshells may become deformed, resulting in complications of the chick's use of shell calcium that in turn makes control of moisture loss difficult.
- Vitamin E The source of this is in seed germs i.e. wheat. A deficiency results in poor hatches of chicks that are weak and do not thrive. Most deaths occur due to circulatory failure in which the walls of the blood vessels are ruptured.
- Vitamin K This can be found in most green feed. A gross deficiency will cause haemorrhages over the body of the chicks. In eggs, deaths may occur in turning due to severe bumping or sudden jarring.
- Vitamins in the B Group
 Vitamin B1 *Thiamine* Vitamin B2 *Riboflavin*
- Nicotinic Acid or Niacin
- Vitamin B6 Pyridoxine

- · Pantothenic Acid
- Biotin
- Folic Acid
- Vitamin B12
- Minerals

- Calcium
- Phosphorous
- Manganese
- Magnesium
- Zinc

2.3. Storage and Handling of Eggs

The quality of the chick all depends on the quality of the egg. It is essential that care is taken in the storage and handling of the eggs. It must be emphasised that the eggs cannot be improved once laid, and will only deteriorate with poor storage and handling. The following factors will help to ensure an egg is of best possible quality:-

NOTE:- Eggs should be stored with pointed end down.

• Temperature

Ideal storage temperature is between 12°C and 15°C (50 – 69°F). Too high a storage temperature and some incubation can commence but at the wrong rate. This will decrease the chances of the embryo developing normally. If storage temperature is below freezing the cell structure of the egg can break down and prevent the embryo developing.

2.3. Storage and Handling of Eggs (continued)

• Humidity

The best humidity level at which to store eggs is between 75 to 85% RH to avoid significant drying out of the egg before incubation.

• Time in Storage

Depending on species, it is preferable that eggs should not be kept for more than 7 days prior to incubation. Between 7 and 14 days hatchability begins to decrease considerably. Vitamins decay and membranes breakdown in time and so the embryo can often suffer early mortality.

Cleanliness

Cracked, mis-shaped and heavily soiled eggs should be discarded (if possible). Soiled eggs may be cleaned using a brand egg wash solution such as Brinsea Incubation Disinfectant Concentrate following the manufacturers instructions. It is essential to wash eggs in solution that is significantly warmer than the egg so that expansion in the egg causes flow out through the pores rather than dirty water flowing inwards. Bear in mind that all solutions will remove the outer cuticle from the egg as well as the dirt and may leave the egg at greater risk from bacterial contamination in the future.

• Turning During Storage

Eggs should be turned once a day, 45 degrees each way, back and forth through 90° during the storage period. Insufficient turning can cause the yolk to float and touch membranes near the shell. If the embryo touches then it may stick and prevent growth once inside the incubator.

• Handle Eggs Carefully

Eggs should be handled with care as severe bumping may rupture the yolk membranes, even if the shell remains undamaged. Reasonable care should also be taken during incubation also. A bump can rupture blood vessels causing the chick to bleed to death.

• Collection of Eggs

Generally the best time to collect eggs is before 9am, and to check again at lunchtime. Eggs left in the nest for much longer than 24 hours tend to be poor hatchers.

• Egg Shape, Shell Texture and Quality

Some naturally misshaped eggs have lower than expected hatch rates. Undersize eggs often have a large yolk in proportion to the albumen. Large eggs can sometimes be 'double yolkers' which will not hatch. Misshaped eggs usually have faults in the shell.

It is important to remember that in natural circumstances eggs warm up quite slowly. Incubators often warm up quickly, and so if placed in the incubator immediately the egg may suffer some thermal shock. If possible, the eggs should be brought up to temperature slowly.

3. Incubator Construction and Function

3.1. Introduction

Incubators are machines, which artificially provide the egg with the correct, controlled environment for the developing chick. Depending on complexity, an incubator will give varying degrees of control over temperature, humidity, egg turning, fresh air flow and hygiene, while providing a secure place for the eggs.

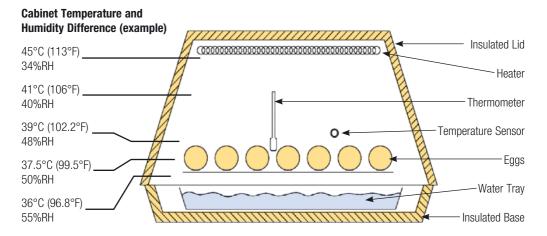
Incubators may be classed as Still Air, Forced Draft and Contact types according to how air is circulated. Contact incubators are a new generation of machines designed to mimic natural incubation much more closely.

3.2. Still Air Incubators

Still air incubators are the most basic form of incubator. A still air incubator is basically an insulated box consisting of:-

- A Heating element
- · A Thermostat or temperature controller to control temperature
- Egg tray
- A thermometer to measure the air temperature
- A tray for water
- Some machines may have an hygrometer for humidity measurement
- · Some machines may have turning mechanism for automatic turning of eggs

The air inside a still air incubator is circulated by convection. As the air is heated it expands and rises to the top of the incubator. The amount of airflow achieved in a still air machine is therefore determined by the ratio of air temperature inside the box to outside. The lower the air temperature outside the box the greater the airflow inside. To achieve good air circulation, air inlets are usually positioned in the base and top of the incubator. Inside a still air incubator, the warm air moves towards the top so different temperatures will be recorded at different levels (see example diagram below). It is therefore important that a still air incubator is kept on a level surface and that eggs are all of similar size.



3.3. Forced Draft Incubator

The forced draft machine was developed to overcome temperature gradient problems throughout the incubator. In a forced draft incubator a fan is used to circulate the air, which gives a uniform temperature throughout the machine. The air temperature surrounding the egg is therefore constant and positioning of the thermometer and temperature sensor are less critical. Eggs, too, can be of differing size and set in trays at different levels.



3.4. Contact Incubators

Contact Incubation mimics the natural incubation process that occurs in the nest.

The most striking difference between natural and artificial incubation is the fact that the natural parent provides warmth by contact rather than surrounding the egg with warm air. This may not at first sight seem significant but important research studies carried out by J. Scott Turner at New York State University and the University of Cape Town has revealed major unsuspected differences in thermal behaviour of eggs incubated by contact rather than convection. Crucially, eggs in a forced draft incubator have near uniform temperature throughout incubation; there will be some increase in embryonic temperature towards the end of incubation as a result of increased metabolism but otherwise the whole egg will remain near incubator air temperature. By contrast, eggs incubated naturally or by contact, have significant temperature differences. Heat is entering the egg over a relatively small brood patch, which is therefore warmer than other regions of the egg and is being lost from most of the

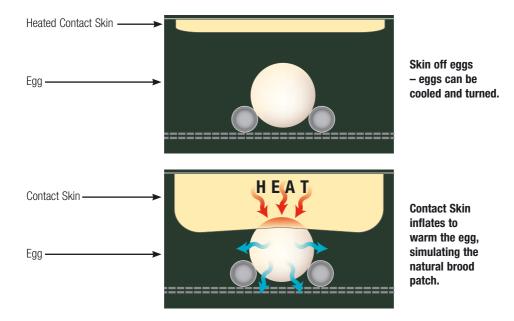
remaining shell area, which is cooler.

In **contact incubation**, the embryo temperature tends to fall at later stages of incubation as a result of the embryo's own blood circulation which becomes significantly more important than embryonic metabolism in determining temperature distribution and heat flow within the egg, quite contrary to previous understanding which assumed metabolism to be dominant and cause egg temperature to rise. The inevitable fact that the embryo grows larger and must move from its original position on top of the yolk sack downwards in the egg to cooler regions also tends to reduce embryonic temperature as incubation progresses. These important findings emphasise surprising differences between natural and artificial incubation, but there are others.

Eggs in nature are certainly subjected to a cycle of warming and cooling coupled with ventilation as the parent bird leaves the nest to feed and defend territory, etc.

The first manufactured contact incubator, Brinsea's 'Contaq' incubator should be viewed as replacing the natural Bird and Nest combination. The lid with the attached 'skin' mimics the functions of an incubating parent; providing warmth by contact with the tops of eggs, but also the facility to lift from the eggs periodically and in doing so, causes a substantial influx of fresh air in the manner of a bird standing or leaving the nest. The skin is gently but firmly pressed against the eggs by a low positive pressure of air, ensuring good thermal conduction.

The egg chamber substitutes for the 'nest' in providing a safe, protected environment for the eggs, with provision for air to be induced through the nest material which enables the operator to exercise control over the degree of ventilation of eggs. Nests of different species have very different characteristics, particularly with respect to gas permeability. This in turn affects water loss from eggs, so it is necessary to adjust the amount of nest material accordingly. Where required, additional humidity is introduced to the egg chamber automatically. The ideal combination of nest material and humidity setting is best determined by weighing eggs and monitoring water loss. The egg chamber is not directly heated.



The Contaq egg chamber also provides sophisticated provision for automatic turning of eggs of different sizes. Egg turning coincides with lifting of the skin and enables programmable fixed or random turns.



3.5. Design Requirements for Successful Hatching

Temperature

Accurate incubation temperature is by far the most important requirement for successful hatching of chicks. Even marginal temperature differences can affect hatch rates, although these differences seem to have less effect on eggs in contact incubators. The growth processes in the development of the embryo are very temperature sensitive and small deviation can cause development to progress out of sequence resulting in losses or deformities. The design basis for Brinsea incubators is a maximum $\pm 0.25C$ across the egg tray and $\pm 0.25C$ over a period of time.

It is the egg centre or embryo temperature, that is most critical. To ensure this is correct it is of highest importance that the incubator is set up correctly, particularly for still air machines. Still air incubators have temperature variations from top to bottom, therefore the sensor of the temperature controller and the thermometer bulb need to be positioned as close as possible to the top of the eggs. Here the temperature needs to be slightly higher than the mean temperature used in forced draft incubators and accounts for the important difference found in the instructions. It is therefore vital to follow the manufacturer's recommendations for the incubator used.

Humidity

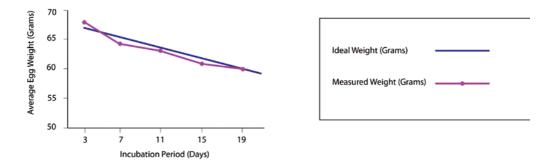
Constant accuracy of humidity is less critical than that of temperature. Ideally, the egg needs to lose 13 - 15% of its weight between the time of laying and pipping, (more for some altricial species). Fairly wide tolerances in humidity are bearable although not ideal, as long as the chick ends up having lost the correct amount of weight by the time of hatching. Correction can be made in later stages for errors earlier.

Experience has shown Contact Incubation to be less critical of exact humidity control, perhaps due to the egg being in a more natural air-flow environment. Highly accurate digital humidity control is available for all Brinsea incubators.

The weight loss rate will vary according the type of egg, the amount of ventilation, the handling of the egg, the breeder diet and the time in the season in which it was hatched. There are two guides that help indicate correct humidity.

Firstly, the air cell. The air cell increases as the incubation proceeds. In order to see this at first sight, the eggs need to be candled (Chapter 4).

The other indicator is weight loss. If you weigh eggs before setting and weigh them as incubation progresses, the weight loss can be plotted on a graph (example below) to determine if the average weight loss has been correct.



Humidity can be adjusted during the incubation period according to the graph reading at that particular time in the incubation process. As a general rule, if actual weights are lower than ideal, then humidity needs to be increased. If actual weights are higher than ideal, then humidity needs to be decreased.

All incubators should have the facility to adjust humidity levels. There are two controllable factors that influence humidity levels. These are the amount of water surface area, and the amount of fresh air that the incubator draws in. The greater the water surface area, and the less fresh air being drawn in, the higher the humidity levels inside the incubator will be. One method to increase water surface area is to use evaporating pads or blotting paper. Finally, the environment in which the incubator is set up in can have an effect on accurate humidity control. If the ambient humidity in the air outside the incubator is very dry, then incubation humidity levels will be lower than if the air is very humid (wet). Also, cold air cannot retain much water vapour, so when cold winter air is warmed the RH level will be very low. This happens in heated houses in winter, and also inside incubators. The result of this in general is that humidity levels tend to be lower in the winter than in summer and so humidity levels should be adjusted with this in mind.

Some breeders go to great lengths to control the incubation room environment and overcome seasonal variations in ambient humidity. In extreme cases, sections of the egg's shell are removed to allow extra water loss, or covered with tape to reduce water loss. This should be regarded as strictly for the experts though and a high risk method.

The humidity levels required when a chick is hatching need to be higher than previously in the incubation period. For the last day or so, high humidity levels are required to prevent the membranes of the egg drying too fast as the chick hatches and becoming tough and difficult to tear. The humidity level when hatching should therefore be at least 60%RH.

Turning

As the embryo develops on the yolk, it causes that part of the yolk to become lighter and float upwards. If, after the egg is moved, the embryo is downwards it will cause the yolk to rotate in the inner thin white until it is uppermost again.

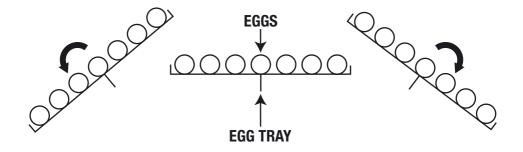
The yolk is held in position by a structure called the chalaza and by membranes. It still tends to float upwards, deforming the inner membranes and eventually the yolk will touch the shell membranes. If this happens with the embryo uppermost the embryo will stick to the shell and die. For this reason, turning is essential for survival of a healthy chick.

As the egg is turned the embryo is swept into fresh nutrients, allowing the embryo to develop. This is critical for the first week when the embryo has no circulation system. After the first week, eggs still need to be turned but not as often. The turning regime is often different between species and altricial birds tend to need more frequent turning than precocial birds. Whereas fowl are turned through 80 degrees every hour or so, parrots are often turned through 180 degrees many times an hour in the early stage.

In incubators today there are four types of turning mechanisms:-

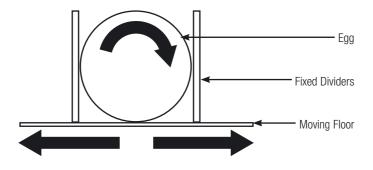
i) Tilting Trays

Most commercial incubators are provided with plastic egg trays that hold the egg vertically, with the small end down. The tray is then tilted through an angle of about 40° either side of horizontal (an overall angle of 80°) at predetermined intervals, perhaps every hour for example. This method works well with poultry for which it was developed, and is very efficient to operate on a commercial scale. However this is very different from the natural process adopted by birds. So, this technique is fine for poultry but not so good for wild or altricial birds.



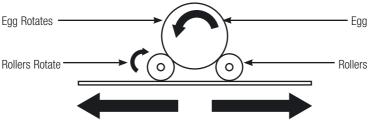
ii) Moving Floor

In some smaller incubators where there is no need for multiple layers of eggs, the eggs lie on a horizontal floor or conveyor, which moves from side to side. Eggs are restrained from the lateral movement by fixed dividers so that the eggs roll. This kind of system works well with eggs which are reasonably symmetrical end for end, for example duck eggs, but pointed eggs tend to bunch together with this system.



iii) Rollers

Rollers work by being rotated by a moving floor. Eggs sit on rollers that in turn sit on the moving floor. Ribs on the rollers help reduce the tendency for an egg to 'walk' along the roller length.



Floor or Conveyor moves

iv) Troughs or Channels

Semicircular shaped tilting channels hold eggs in position as they are turned preventing eggs 'creeping' as can happen with rollers or moving floor. Eggs up to a maximum size and be safely turned, determined by the channel dimensions. Shown in the Brinsea Ovation 28.



4. Monitoring of Eggs and the Incubator

Eggs need to be monitored during incubation to determine embryo progress and where incubation adjustment may be necessary.

4.1. Candling

Candling lamps are lights with a concentrated beam that may be shone through the shell of the egg to illuminate the egg contents. This allows the size of the airspace to be determined which offers a guide to the weight loss rate. If the air space is larger than expected too much water is being lost and the humidity in the incubator should be increased to reduce the rate of water loss. If the air space is smaller than expected then the opposite applies.

Embryos can be damaged by the heat from conventional candling lamps which is why Brinsea's OvaView candling lamps use high power yet cool LEDs to prevent this problem and still be highly effective. The matching OvaScope shrouds the candled egg making visibility of the embryo even better and means that the user doesn't need to dim the room light which is deal for classrooms

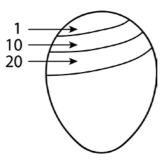
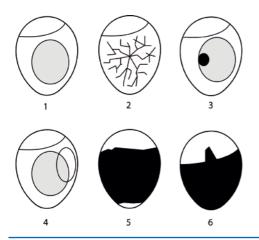


Diagram shows extent of airspace development throughout incubation (in days)

Candling also allows the development to be observed so that eggs that are infertile or have died may be safely removed from the incubator.



- **1.** Clear when candled probably infertile (or very early death) when candled at 1/3 of the incubation period.
- 2. Fertile with red blood vessels after 1/3 of the incubation period.
- **3.** Red or black staining early death when candled after 1/3 of the incubation period.
- **4.** Embryo with red blood 'ring' early death when candled after 1/3 of the incubation period.
- **5.** Dark outline with ill defined detail late death (1/2 or 2/3 through incubation period).
- **6.** Live embryo with bill in air sack due to hatch in 24-48 hours.

4.2. Measuring Humidity and Weight Loss

Modern sensor technology allows direct measurement of humidity although accuracy of these devices varies, with much higher accuracy sensor adding to incubator cost. Avoid mechanical dial hygrometers as these are often inaccurate. Brinsea only use high accuracy humidity measuring systems as we believe an inaccurate measurement is worse than no measurement at all.

Weight Loss Calculations

If the weight of an egg was not measured when it was laid it may be estimated using the formula:

Fresh Laid Weight (g) = Length (cm) x Breadth (cm) x Breadth (cm) x 0.548

Using the fresh laid weight and the weight loss percentage recommended for the species the daily weight loss may be calculated:

 Daily Weight Loss =
 Fresh Laid Weight x Desired % Loss to pip

 Number of Days to Pip

Some breeders prefer to use change of Egg Density as the measure of water loss from the egg:

Egg Volume = Length (cm) x Breadth (cm) x Breadth (cm) x 0.51

Egg Density $(g/cm^3) =$

Egg Weight (g) Egg Volume (cm³)

Estimated Daily Change in Egg Density:

Daily Change =

Egg Density at Time (T1) – Egg Density at Time (T2)

Time (T2) – Time (T1)

5. Chick brooders

Once chicks have dried out they need to be removed from the incubator but still kept warm. Precocial species (poultry, waterfowl and game birds) are mobile and so just need a safe environment, appropriate food and water and a source of heat and they will look after themselves. Altricial species, hatching blind and unable to move, need more intensive brooding and are usually placed in a temperature controlled intensive care brooder (such as Brinsea's TLC-40) where the temperature is controlled and can be gradually reduced as the chicks grow.

It's never a good idea for chicks to be in constant light, especially for game bird chicks where 24 light can stress the chick leading to them attacking each other. Brinsea's Ecoglow models are ideal for brooding precocial species as they emit no light, are very efficient and operate from 12 volts, eliminating all electrical safety problems.'



Brinsea EcoGlow Safety 1200

6. Troubleshooting

| Problem | Possible Causes |
|---|--|
| Chicks hatch late | Large eggs Old breeders Eggs stored for too long Incubator temperature too low Weak embryos Inbreeding Incubator humidity too high |
| Slow (drawn-out) hatch | Mix of eggs (different sizes, different aged breeders, storage times) Poor egg handling Hot or cold spots in incubator Incubator or Hatcher temperature too high or low |
| Sticky Chicks, smeared with albumen | Low incubation temperature High incubation humidity Poor turning Old eggs Very large eggs |
| Chicks stuck in shell, dry, shell fragment stuck to feathers | Low humidity in storage Poor egg turning Cracked shell or Poor shell quality |
| Premature hatching, bloody navels | • Temperature too high |
| Small chicks | Small eggs Low humidity High temperature High altitude Thin, porous shells |

| Problem | Possible Causes |
|---|--|
| Unhealed naval, dry, rough down feathers | High incubator temperature, or temperature fluctuations Humidity too high when hatching Inadequate nutrition |
| Weak Chicks | High hatching temperaturePoor Hatcher ventilationContamination |
| Chicks malpositioned | Eggs sat small end up position Inadequate turning Excessive turning at late stages Too high or too low temperature High humidity Old breeders Round shaped eggs Nutritional deficiencies Retarded development Poor egg handling or storage conditions |
| Malformations | Poor storage conditions Jarring of eggs Nutritional deficiencies Inadequate turning High or low temperature Inadequate ventilation |
| Crooked toes, bent legs | High or low temperaturePoor nutrition |
| Short down, wiry down | Nutritional deficiencies (especially riboflavin) High incubation temperature |

6. Troubleshooting (continued)

| Problem | Possible Causes |
|---|--|
| Eyes closed, down stuck to eyes | Temperature too high in Hatcher Chicks remain in Hatcher too long after hatching Excessive air movement in Hatcher |
| Exploding eggs | Dirty eggs from nest Eggs not washed properly (eggs wiped or cleaned with contaminated cloth) Water condensation on eggs Water sprayed on eggs Contamination from earlier exploders Contaminations from handling with dirty hands |
| Dwarf embryos, runts in growing chicks | Egg contaminationsBreeder diseasesNutritional deficiencies |
| Haemorrhage | Incubator or Hatcher temperature too high Rough handling at transfer Nutritional deficiencies (vitamin K or vitamin E) Contamination |
| Swollen head and back of neck | Nutritional deficiencies |
| Small air cell, egg weight loss under 10% | High humidityVery thick shellsLow temperature |
| Exposed brain | High incubation temperature Low oxygen levels |

Incubation Reference Table

| Species of Egg | Incubation Period | Still / Forced Air Incubation | | | | |
|----------------------------|----------------------|----------------------------------|-----------------|------------------------|-----------------|--|
| | (Days) | Temperature °C (°F) | Humidity RH% | Temperature °C (°F) | Humidity RH% | |
| Hens | 21 | 37.5 (99.5) | 40 — 50 | 39.5 (103.1) | 40 – 50 | |
| Pheasant | 23 – 27 | 37.3 (99.1) | 40 - 50 | 39.7 (103.5) | 40 - 50 | |
| Partridge | 23 – 24 | 37.5 (99.5) | 40 - 50 | 39.5 (103.1) | 40 - 50 | |
| Quail | 16 – 23 | 37.3 (99.1) | 40 - 50 | 39.7 (103.5) | 40 - 50 | |
| Ducks | 28 | 37.5 (99.5) | 45 – 55 | 39.5 (103.1) | 45 – 55 | |
| Geese | 28 – 32 | 37.3 (99.5) | 45 – 55 | 39.5 (103.1) | 45 – 55 | |
| Falcons | 31 – 33 | 37.0 (98.6) | 40 – 45 | 39.0 (102.2) | 40 - 45 | |
| Merlins | 28 – 32 | 37.5 (99.5) | 50 | 39.5 (103.1) | 50 | |
| Kestrels | 27 – 29 | | 50 | | 50 | |
| Barn Owl | 30 | | 50 | | 50 | |
| Tawny Owl | 29 | | 50 | | 50 | |
| Snowy Owl | 33 | | 50 | | 50 | |
| Amazons (Parrots) | 24 – 29 | 37.3 (99.1) | 35 – 45 | 39.0 (102.2) | 35 – 45 | |
| Macaws (Parrots) | 28 – 30 | 37.3 (99.1) | 35 – 45 | 39.0 (102.2) | 35 – 45 | |
| Love Birds (Parrots) | 22 – 24 | 37.3 (99.1) | 35 – 45 | 39.0 (102.2) | 35 – 45 | |
| African Grey (Parrots) | 26 – 28 | 37.3 (99.1) | 35 – 45 | 39.0 (102.2) | 35 – 45 | |
| Sulphur Cockatoo (Parrots) | 29 – 31 | 37.3 (99.1) | 35 – 45 | 39.0 (102.2) | 35 – 45 | |
| Electus (Parrots) | 28 | 37.3 (99.1) | 35 – 45 | 39.0 (102.2) | 35 – 45 | |
| Rhea | 35 - 40 | 36.0 (96.8) | 20 – 25 | 38.0 (100.4) | 20 – 25 | |
| Emu | 49 – 52 | 37.0 (98.6) | 20 – 25 | 39.0 (102.2) | 20 – 25 | |
| Ostrich | 40 - 43 | 36.4 (97.5) | 20 – 40 | 38.4 (101.0) | 20 - 40 | |

IMPORTANT:-

Temperatures for Still Air incubators MUST be adjusted in accordance with the Manufacturer's instructions to take account of the difference in temperature at different levels in the machine.

While great effort is taken to ensure the accuracy of data, no responsibility can be accepted for any loss or injury incurred due to error or omission. Figures given are guidelines only and some variation may be experienced in practice.

NOTES

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